

## CRISmono™ IDH1 Quick Guide

### Preparation for Assay

- Before starting, spin down all reagents briefly to collect components clinging under the lid and keep them on ice until ready for use.
- In the case of frozen reagents, they are fully thawed and mixed using a vortex mixer.
- Briefly spin down and keep on ice until ready for use.
- Using a filter tip during all experimental processes is recommended.

### Step 1. IVC (Total volume : 10 µL, each)

1-1) Prepare a reaction mix by adding the reagents in the order indicated in the following table to a clean PCR tube:

| Reagent                 | Volume per Sample |
|-------------------------|-------------------|
| ● Remov RXN buffer      | 4 µL              |
| ● Stabilizer            | 1 µL              |
| ● IDH1 Enzyme mix       | 4 µL              |
| Genomic DNA (20–150 ng) | 1 µL to each tube |
| Total Volume 10 µL      |                   |

1-2) Mix each reagent, then perform vortexing and spin down.

1-3) Place the PCR tube in the pre-set thermal cycler under the conditions shown in the table below and run the program.  
(Lid temperature: 60 °C)

| Step description               | Temperature | Time   |
|--------------------------------|-------------|--------|
| <i>In vitro</i> Cleavage (IVC) | 45 °C       | 60 min |

1-4) After the 60 min reaction, add ● 1 µL of 10X STOP buffer to each reaction mixture and mix by pipetting or vortex.

### Step 2. Target amplification (Total volume : 50 µL, each)

2-1) Prepare a reaction mix by adding the reagents listed in the following table to a new PCR tube:

| Reagent             | Volume per Sample |
|---------------------|-------------------|
| ● IDH1 Primer mix   | 5 µL              |
| ○ 2X PCR Master mix | 25 µL             |
| Nuclease-free Water | 18 µL             |
| IVC product         | 2 µL              |
| Total Volume 50 µL  |                   |

2-2) Mix each reagent, then perform vortexing and spin down.

2-3) Place the PCR tube in the pre-set thermal cycler under the conditions shown in the table below and run the program.  
(Lid temperature: 105 °C)

| Step description | Temperature | Time   | Cycles |
|------------------|-------------|--------|--------|
| Pre-denaturation | 98 °C       | 3 min  | 1      |
| Denaturation     | 98 °C       | 10 sec | 42     |
| Annealing        | 55 °C       | 40 sec |        |
| Extension        | 72 °C       | 30 sec |        |
| Final Extension  | 72 °C       | 5 min  | 1      |
| Hold             | 4 °C        | ∞      |        |

### Step 3. DNA Purification (Clean up)

3-1) Purify DNA from the reaction mixture.

: Column purification, magnetic beads size selection and enzyme purification are possible to use

### Step 4. Sanger sequencing

4-1) Please refer to the manufacturer's instructions and recommendations.

#### Reagent

- IDH1 Seq Primer F (10 pmole/µL)

### Step 5. Data analysis

5-1) Sequencing chromatogram analysis for IDH1 codon 132 (CDS 394–396) sites.

: TGGGTA AACCTATCATCATAGGT **CGT** CATGCTTATGGG-GATCAAGTAAGT

5-2) VAF (%) is determined by the ratio of peak heights and is considered positive when the VAF value exceeds 10%.

$$\text{VAF (\%)} = \frac{\text{Mutant height}}{\text{Wild-type height} + \text{Mutant height}} \times 100$$